

Patients with KCNH1 -related intellectual disability without distinctive features of Zimmermann-Laband/Temple-Baraitser syndrome

Marion Aubert Mucca, Olivier Patat, Sandra Whalen, Lionel Arnaud, Giulia Barcia, Julien Buratti, Benjamin Cogné, Diane Doummar, Caroline Karsenty,

Sandra Kenis, et al.

► To cite this version:

Marion Aubert Mucca, Olivier Patat, Sandra Whalen, Lionel Arnaud, Giulia Barcia, et al.. Patients with KCNH1 -related intellectual disability without distinctive features of Zimmermann-Laband/Temple-Baraitser syndrome. Journal of Medical Genetics, 2021, pp.jmedgenet-2020-107511. 10.1136/jmedgenet-2020-107511. hal-03244899

HAL Id: hal-03244899 https://u-paris.hal.science/hal-03244899v1

Submitted on 15 Mar 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



This item is the archived peer-reviewed author-version of:

Patients with KCNH1-related intellectual disability without distinctive features of Zimmermann-Laband/Temple-Baraitser syndrome

Reference:

Aubert Mucca Marion, Patat Olivier, Whalen Sandra, Arnaud Lionel, Barcia Giulia, Buratti Julien, Cogné Benjamin, Doummar Diane, Karsenty Caroline, Kenis Sandra,- Patients with KCNH1-related intellectual disability without distinctive features of Zimmermann-Laband/Temple-Baraitser syndrome Journal of medical genetics - ISSN 1468-6244 - (2021)6 p. Full text (Publisher's DOI): https://doi.org/10.1136/JMEDGENET-2020-107511 To cite this reference: https://hdl.handle.net/10067/1809370151162165141

uantwerpen.be

Institutional repository IRUA

1 Patients with *KCNH1*-related intellectual disability without distinctive fea-

2 tures of Zimmermann-Laband/Temple-Baraitser syndrome

3

4 AUTHORS

Marion Aubert Mucca¹, Olivier Patat^{*1}, Sandra Whalen², Lionel Arnaud³, Giulia
Barcia⁴, Julien Buratti³, Benjamin Cogné⁵, Diane Doummar⁶, Caroline
Karsenty⁷, Sandra Kenis⁸, Eric Le Guern³, Gaëtan Lesca^{9,10}, Caroline Nava³,
Mathilde Nizon⁵, Amélie Piton¹¹, Stéphanie Valence⁶, Laurent Villard¹², Sarah
Weckhuysen¹³, Boris Keren³, Cyril Mignot^{3,10,14,15}

- 10 1. Service de Génétique Médicale, CHU Toulouse, France 11 2. Unité Fonctionnelle de Génétique Clinique, Centre de Référence Anomalies du dé-12 veloppement et syndromes malformatifs, APHP.Sorbonne Université, Hôpital Ar-13 mand Trousseau, France 14 3. APHP.Sorbonne Université, Département de Génétique, Groupe Hospitalier Pitié 15 Salpêtrière et Hôpital Trousseau, Paris, France 16 4. APHP, Service de Génétique Médicale, Hôpital Necker-Enfants Malades, Imagine 17 Institute, Paris Descartes University, Paris, France. 18 5. CHU Nantes, Service de Génétique Médicale, L'institut du thorax, INSERM, CNRS, 19 **UNIV Nantes, Nantes, France** 20 6. Service de Neuropédiatrie - Unité de neuropédiatrie et pathologie du développe-21 ment, CHU Paris Est - Hôpital d'Enfants Armand-Trousseau, France 22 7. Service de Neuropédiatrie, CHU Toulouse, France 23 8. Child Neurology Department, University of Antwerp, Belgium - Neurology Depart-24 ment, University Hospital Antwerp, Belgium 25 9. Service de Génétique - LBMMS, Groupement Hospitalier Est, CHU de Lyon, 26 France 27 10. EuroEPINOMICS 28 11. Laboratory of Genetic Diagnosis, Hôpitaux Universitaires de Strasbourg, Stras-29 bourg, France 30 12. Laboratoire de génétique moléculaire, Département de génétique médicale, CHU 31 de Marseille - Hôpital de la Timone, France 32 13. Applied&Translational Genomics group, VIB-Center for Molecular Neurology, Uni-33 versity of Antwerp, Belgium - Neurology Department, University Hospital Antwerp, 34 Belgium 35 14. Centre de Référence Déficiences Intellectuelles de Causes Rares, Paris, France 36 15. INSERM, U1127, CNRS UMR 7225, Sorbonne Université, UPMC Univ Paris 06 37 UMR S 1127, Institut du Cerveau et de la Moelle épinière, ICM 38 39 ***CORRESPONDING AUTHOR** 40 Dr Olivier Patat, Service de Génétique médicale, Hôpital Purpan, CHU TOULOUSE, 41 Place du Dr Baylac, 31300 TOULOUSE 42 Email: patat.o@chu-toulouse.fr
- 43 WORD COUNT: text 1997 words; abstract 200 words

44 ABSTRACT

De novo missense variants in *KCNH1* encoding Kv10.1 are responsible for two clinically recognizable phenotypes: Temple-Baraitser Syndrome (TBS) and Zimmermann-Laband Syndrome (ZLS). The clinical overlap between these two syndromes suggests that they belong to a spectrum of *KCNH1*-related encephalopathies. Affected patients have severe intellectual disability (ID) with or without epilepsy, hypertrichosis and distinctive features such as gingival hyperplasia and nail hypo/aplasia (present in 20/23 reported cases).

52 We report a series of seven patients with ID and de novo pathogenic KCNH1 53 variants identified by whole-exome sequencing or an epilepsy gene panel in 54 whom the diagnosis of TBS/ZLS had not been first considered. Four of these 55 variants, p.(Thr294Met), p.(Ala492Asp), p.(Thr493Asn) and p.(Gly496Arg), were located in the transmembrane domains S3 and S6 of Kv10.1 and one, 56 57 p.(Arg693Gln), in its C-terminal cyclic nucleotide binding homology domain (CNBHD). Clinical reappraisal by the referring clinical geneticists confirmed the 58 59 absence of the distinctive gingival and nail features of TBS/ZLS.

60 Our study expands the phenotypical spectrum of *KCNH1*-related encephalopa-61 thies to individuals with an attenuated extra-neurological phenotype preventing 62 a clinical diagnosis of TBS or ZLS. This subtype may be related to recurrent 63 substitutions of the Gly496, suggesting a genotype-phenotype correlation and, 64 possibly, to variants in the CNBHD domain.

65 **KEYWORDS**

66 *KCNH1*, intellectual disability, epilepsy, Temple-Baraitser syndrome, Zimmer-67 mann-Laband syndrome

68 **INTRODUCTION**

69 Gain-of-function missense variants in KCNH1 (MIM*603305) encoding the volt-70 age gated potassium channel Kv10.1 are associated with syndromic neurode-71 velopmental disorders with overlapping phenotypes referred to as Temple-72 Baraitser syndrome (TBS;MIM#611816) and Zimmermann-Laband syndrome 73 (ZLS;MIM#135500).[1-7] The main features of these syndromes (Table S1) are 74 severe intellectual disability (ID) with or without early-onset epilepsy, congenital 75 hypertrichosis, anomalies of the limbs and nails, gingival hyperplasia and mild craniofacial dysmorphic features.[1-3, 7-13] Gingival hyperplasia is a classical 76 77 feature of the ZLS phenotype, present in about 50% of the published cases of 78 KCNH1-related encephalopathy.[11-14] In patients with TBS, thumbs and hal-79 luces are usually long, proximally implanted and broad with hypoplastic nails 80 and an increased radiolucency with missing ossification points in the terminal 81 phalanx.[5, 15] The other fingers and toes may show hypoplastic distal phalan-82 ges and nails.[10] Hypo/aplasia of nails has been reported in more than 90% of 83 published cases with KCNH1 variants.[4, 5, 7, 15, 16] These findings in a con-84 text of severe encephalopathy are highly suggestive of TBS/ZLS.

We report on seven novel patients with *KCNH1*-related ID and lacking the distinctive features of TBS/ZLS.

87

88 METHODS

We used clinical networks (EPIGENE, EuroEPINOMICS-RES, GeneMatcher) to collect a multicentric cohort of patients with *KCNH1*-related encephalopathy, evaluated by clinical geneticists for the etiological diagnosis of ID. Referring 92 physicians retrospectively completed a standardized table with molecular and 93 clinical data. We obtained written consents for genetic testing and publication of 94 data for all patients. Retrospectively, craniofacial features, limbs and nails were 95 assessed using patients' photographs.

96 Patients P1, P4 and P5 were studied on epilepsy gene panels used in different 97 French University hospitals, including 90 to 116 genes involved in epilepsy, 98 among 2450 individuals tested in different French university hospitals. Patients 99 P2, P3, P6 and their parents were studied by solo or trio-WES (supplementary 100 methods). The variant found in patient P2 was identified by performing WES in 101 a research project on genetic ataxias. The variants in patients P3 and P6 were 102 found by WES in the diagnosis workup of ID out of 1294 analyzed patients. Pa-103 tient P7 was recruited through GeneMatcher and the KCNH1 variant was found 104 by WES (1/850 tested patients).

105 Sanger sequencing was used for confirming variants.

Sequence variations were numbered using the adenine of the ATG initiation
codon as the first nucleotide (*KCNH1* GenBank accession NM_172362).

108 We performed an exhaustive review of the literature using the PubMed data-109 base to compile clinical data on individuals with pathogenic variants in KCNH1 110 (Table S1) using the following terms: « KCNH1, TBS, ZLS, Temple-Baraitser, 111 Zimmermann-Laband ». Because genetic heterogeneity has been 112 documented,[13, 17] we excluded patients without mutation to focus on the fre-113 quency of clinical signs in proven KCNH1-related encephalopathy.

INICAL DATA OF OUR 7 PATIENTS WITH KCNH1-RELATED ENCEPHALOPATHY

	P2	P3	P4	P5	P6	P7	ΤΟΤΑΙ
	F	Μ	F	F	F	F	n=7
i y verian	15 y Furopean	7.9 y Mauritian	9 y Furopean	1.75 y European	3.5 y European	28 y European	1.75 y
210977496G>T	g.210977485C>T	g.210948724C>T	g.210977485C>T	g.210977485C>T	g.210977493G>T	g.211192276G>A	
475C>A	c.1486G>A	c.2078G>A	c.1486G>A	c.1486G>A	c.1478C>A	c.881C>T	
Ala492Asp)	p.(Gly496Arg)	p.(Arg693Gln)	p.(Gly496Arg)	p.(Gly496Arg)	p.(Thr493Asn)	p.(Thr294Met)	
PS2 PM1 PM2 '2 PP3)	5 (PS2 PM1 PM2 PP2 PP3 PP5) 0	5 (PS2 PM1 PM2 PP2 PP3) 0	5 (PS2 PM1 PM2 PP2 PP3 PP5) 0	5 (PS2 PM1 PM2 PP2 PP3 PP5) 0	5 (PS2 PM1 PM2 PP2 PP3) 0	5 (PS2 PM1 PM2 PP2 PP3) 0	
.2	29.9 de novo	26 de novo	29.9 likely de novo*	29.9 de novo	26.7 de novo	25.7 de novo	
vere ID	moderate ID	severe ID	mild/moderate ID	mild DD	moderate/severe ID	severe ID	mild -
m	10 m	10 m	12 m	9.5 m	26 m	NA	9.5 m -
t achieved	21 m	35 m	20 m	18 m	not achieved	18 m	18 m -
calization	sentences	few words	sentences	canonical babbling	no words	no words	4.5 y senter words
S	yes	no	yes	yes	no	no	4/7
h	12 m	NA	2 у	3 m	NA	NA	day 1 -
ıic-clonic, myo- ınic	tonic, tonic-clonic; 2 status epilepticus	NA	focal clonic, gener- alized tonic-clonic (total 5 seizures), myoclonic	generalized tonic- clonic, focal clonic, myoclonic	NA	NA	
veek	monthly/weekly	NA	1-3/month	8/day	NA	NA	
w background tivity, focus of left ntal spike-and- ve	normal	NA	focus of central spikes increased during sleep (close to CSWS)	few spikes in cen- tral regions, bursts of generalized polyspikes with myoclonic jerks	slow background activity	normal	
sponsive	responsive	NA	responsive	responsive with	NA	NA	
retiracetam	lamotrigine, carba- mazepine and clobazam	NA	carbamazepine	levetiracetam	NA	NA	
rmal	normal	normal	normal	normal	normal	normal	norma 7/7
oderate sensori- ural deafness, opia and asti-	hyperopia	none	none	none	none	none	
tism ncal and periph- al hypotonia	truncal hypotonia, hyperreflexia, mild ataxia	hyperreflexia	NA	normal	truncal hypotonia	facial hypotonia, hyperreflexia	

1 PATIENTS AND RESULTS

2 We collected clinical and molecular data of seven unrelated patients with path-3 ogenic *KCNH1* variants (Table 1).

4 Three individuals shared the same previously reported *de novo* (or assumed as de novo) heterozygous missense variant c.1486G>A, p.(Gly496Arg). The others 5 6 carried novel de novo heterozygous missense variants [c.881C>T p.(Thr294Met), c.1475C>A p.(Ala492Asp), c.2078G>A p.(Arg693Gln) and 7 8 c.1478C>A p.(Thr493Asn)] fulfilling the criteria of pathogenicity (Figure 1A, Ta-9 ble1).

10 All seven patients, aged 21 months to 28 years, presented with mild/moderate 11 to severe developmental delay (DD) or ID (Table1). The age of independent 12 walking was mildly delayed (18 to 35 months) for five patients, while the others 13 had not achieved walking at last examination (3.5 and 4.5 years). Two out of the 14 five patients older than 3 years were able to make sentences, one spoke a few words and the remaining three did not speak. Four patients had epilepsy start-15 16 ing in infancy, with generalized tonic-clonic (4/4), myoclonic (2/4), focal motor 17 (2/4) and tonic seizures (1/4). One of them experienced status epilepticus. The 18 epilepsy was pharmaco-responsive in all individuals: a monotherapy was effec-19 tive in three patients (2/3 levetiracetam, 1/3 carbamazepine) and one individual 20 needed a bitherapy (carbamazepine and clobazam). Cerebral MRI was normal 21 in all cases. P1 had a moderate sensorineural deafness.

At examination, the most frequent neurological sign was truncal hypotonia, brisk tendon reflexes and ataxia. The mild dysmorphic features noted in some of the patients were regarded as nonspecific by the physicians and did not guide the molecular diagnosis. Referring clinicians reassessed the phenotypes of their patients after the identification of the *KCNH1* variant with special attention to the morphology of limbs and gums (Figure S1). Only hypertrichosis of the back was present in P1. Almost all patients had constipation (6/7), sometimes severe, and near half of them (3/7) had gastroesophageal reflux disease (GERD).

30

31 **DISCUSSION**

We provide a clinical description of seven individuals with *KCNH1*-related encephalopathy without features of TBS/ZLS found by WES or with an epilepsy gene panel. Only subtle and non-distinctive dysmorphic features were noted in some of them (Figure S1). Only one patient had mild toenails hypoplasia, but this was inherited from his mother and maternal grandfather. Three patients were reported with slightly broad halluces, two with proximal placement of thumbs, including one having long fingers.

39 Twenty-three patients with KCNH1-related encephalopathy have been reported 40 to date (Table S1).[4-9, 13-16, 18, 19] Most of them had a clinical diagnosis of 41 TBS/ZLS based on distinctive clinical features prior to molecular analysis. Two 42 published series reported eight patients with pathogenic KCNH1 variants found 43 by WES[4, 16] without prior clinical diagnosis of TBS or ZLS, but all had gingival 44 hyperplasia and/or nail anomalies in retrospect, except one who carried the Gly496Glu variant.[16] Thus, 22/23 previously reported patients with KCNH1-45 46 related encephalopathy had a prospective or retrospective diagnosis of TBS/ZLS with either nail hypo/aplasia or gingival hypertrophy. In comparison, 47 48 the patients of the present series stand out for their attenuated extra49 neurological phenotype and the lack of clinical clue suggesting TBS or ZLS. Do
50 these patients have a different developmental or epileptic outcome than
51 TBS/ZLS patients?

52

53 All 23 previously reported patients with KCNH1-related encephalopathy had 54 DD/ID. When mentioned (n=16), the ID was rarely mild (n=1), mild/moderate 55 (n=1), or moderate (n=1), and most frequently severe (n=8), severe/profound 56 (n=2) or profound (n=3). The range of DD/ID in our patients is consistent with 57 the literature (Table 1). Seven out of 14 previously reported patients over 18 58 months achieved walking at a mean age of 3.3 years, while 4/6 of our series 59 walked at a mean age of 2 years. Ten out of 12 previous patients older than 2 60 years had not acquired spoken language. In our series, two individuals out of 61 five spoke words or sentences. Thus, language abilities may be mildly better in 62 the patients of our study.

Epilepsy was frequent in our series (n=4/7) and in the literature (n=19/22). First 63 64 seizures occurred between the first day of life and the age of 2 years (mean age 1 year) in our patients, and at a median age of 0.9 year (mean age 2.5 years) in 65 66 the literature (n=18). Seizures were mainly motor in nature, either focal or gen-67 eralized, including clonic, tonic-clonic, tonic and myoclonic seizures, both in the 68 literature and in our series. EEG revealed unilateral or bilateral spikes or spikes-69 waves in fronto-centro-temporal regions in three of our epileptic patients. In the 70 literature, multifocal epileptiform anomalies, sometimes predominating in anteri-71 or or fronto-temporal regions[15] and foci of spikes in frontal and central areas 72 were both reported.[16] One of our patients experienced convulsive status epiProvide the level of DD/ID and the epilepsy.

Nail anomalies and gingival hypertrophy were reported in 90% and 50% of patients in the literature, respectively (Table S1). The lack of nail anomalies and gingival hyperplasia in our series is noteworthy and suggests an overestimation of their frequency in *KCNH1*-encephalopathy. We cannot exclude, however, the appearance of gingival hyperplasia with time in our patients.

Digestive disorders are a known complication of *KCNH1*-related encephalopathy, constipation and GERD affecting 60% and 25% of the patients respectively (Table S1). Though these comorbidities are not surprising in a context of severe encephalopathy, their severity in some of our patients is noteworthy.

A previous study reported four patients with *KCNH1*-related encephalopathy out of 1447 with ID (2.7‰).[4] We found three French patients out of 2450 studied on an epilepsy gene panel and three by WES out of 2144 with ID (total 6/4594). We conclude that the prevalence of the *KCNH1*-related encephalopathy in individuals with epilepsy/ID and without features of TBS/ZLS in France is about 1.3‰.

93

Three unrelated index cases of our series were heterozygous for the c.1486G>A p.(Gly496Arg) variant. This variant and the c.1487G>A (p.Gly496Glu) substitution affecting the same Glycine residue have already

97 been reported in two patients with KCNH1-related encephalopathy.[7, 15, 16] 98 Interestingly, both patients also had mild extra-neurological phenotypes: the 99 individual carrying p.(Gly496Arg) did not have nail hypo/aplasia (but gingival 100 enlargement and moderate hypertrichosis), [7, 15] and the other with 101 p.(Gly496Glu) only presented broad thumbs/toes but no nail aplasia, hypertri-102 chosis, or gingival enlargement.[16] Since these patients are the only published 103 cases of KCNH1-related encephalopathy lacking these key features of 104 TBS/ZLS, this raises the question of a potential genotype-phenotype correlation 105 associating mild or absent extra-neurological signs and variants affecting the 106 Gly496 residue.

107 Functionally tested variants in KCNH1 usually lead to a left-ward shift in voltage dependent activation, producing an increase in K⁺ conductance in the negative 108 109 potential-range (or a so called gain-of-function effect).[7] These variants may 110 then contribute, as suggested by Kortum et al., to a functional blockade of voltage gated Na+ and L-type Ca²⁺ channels through stabilization of the membrane 111 112 potential to a more negative voltage range, and draw parallels with the gingival 113 hyperplasia that can be seen with prolonged use of the Na+ channel blocker phenotyoin or the Ca²⁺ channel blocker nifedipine.[7] KCNH1 is also required for 114 115 resorption of the primary cilium when the cell cycle is reinitiated: hyperactivation 116 of KCNH1 was indeed shown to increase resorption. Alterations of the sonic hedgehog (SHH) cilia-mediated pathway involved in morphogenesis were there-117 118 fore proposed to lead to the skeletal and nail malformations.[17, 20]

When functionally tested in a heterologous cell system, the homotetrameric 120 p.(Gly496Arg) mutant KCNH1 channel failed to produce depolarization-121 122 activated currents demonstrating a loss of function effect. When the 123 p.(Gly496Arg) mutant was co-expressed with wild-type KCNH1 to produce het-124 erotetrameric channels, an increase in K+ conductance was seen at negative 125 membrane potentials.[7] In addition, similarly to other studied pathogenic vari-126 ants, significant reduced K+ conductance was reported at depolarizing poten-127 tials.[7]

128 It is tempting to hypothesize that the absence or subtlety of extra-neurological 129 symptoms in patients with specific variants such as p.(Gly496Arg) compared to 130 variants associated with a "classical" ZLS/TBS phenotype is reflecting tissue-131 specific sensitivities to a more limited gain-of-function effect of this variant. More 132 in-depth comparative studies of the effect of specific KCNH1 variants on SHH 133 pathway and voltage-gated calcium channel activity is warranted, preferably in 134 model systems recapitulating the human phenotype, such as human induced 135 pluripotent stem cell derived neuronal and chondroprogenitor cells.

136

137

Two of our novel variants, c.1475C>A p.(Ala492Asp) and c.1478C>A p.(Thr493Asn), are located in the S6 transmembrane of Kv10.1 (Figure 1B) where other variants [c.1480A>G p.(Ile494Val) and the c.1465C>T p.(Leu489Phe)] have been previously reported in typical ZLS/TBS patients.[4, 7, 15] The c.881C>T p.(Thr294Met) variant is for its part located in the S3 transmembrane domain. The c.2078G>A p.(Arg693Gln) variant is the first reported to

affect a residue in the C-terminal cyclic nucleotide binding homology domain (CNBHD). The biochemical and electrophysiological consequences of this change have to be determined, but could impact the interaction between the *ether-a-go-go* domain and the CNBHD, leading to an impairment of the activation/deactivation channel control.[21]

To further investigate the potential genotype-phenotype correlations in *KCNH1*related encephalopathy, the description of further thoroughly phenotyped cases will be necessary. Meanwhile, the possibility of non-syndromic *KCNH1*-related encephalopathies should be considered when designing epilepsy gene panels or interpreting whole exome data from individuals with epilepsy or non-specific ID.

156 **ACKNOWLEDGEMENTS**

- 157 We acknowledge generous support from the patients and their families. Several
- authors of this publication are members of the European Reference Network on
- 159 Rare Congenital Malformations and Rare Intellectual Disability ERN-
- 160 ITHACA [EU Framework Partnership Agreement ID: 3HP-HP-FPA ERN-01-
- 161 **2016/739516**].

162 CONFLICT OF INTEREST

163 The authors declare no conflict of interest.

164 FUNDING

165 No financial assistance was received in support of the study.

166 ETHICS APPROVAL

- 167 This study was designed in compliance with the tenets of the Helsinki Declara-
- 168 tion.

169 **PATIENT CONSENT**

- 170 Written consents for genetic testing and publication were obtained locally from
- all participants or their representatives.

172 AUTHOR CONTRIBUTIONS

- 173 MAM/OP: Data collection, Data analysis, Manuscript writing, Manuscript editing
- 174 SWh/SWe: Data collection, Manuscript writing, Manuscript editing
- 175 LA/DD/CK/SK/ELG/GL/CN/SV/LV/BK/BC/MN: Data collection, Manuscript edit-
- 176 ing
- 177 GB/JB/AP: Data collection
- 178 CM: Study conception, Data collection, Data analysis, Manuscript writing, Man-
- 179 uscript editing

180 All authors read and approved the final manuscript.

181 Figure 1. Schematic representation of all (likely) pathogenic variants iden-

- 182 tified in *KCNH1*.
- 183 **A. cDNA level**: Legend: in bold, the novel variants reported here.
- 184 **B. protein level:** Legend: Stars representing variants identified, in blue = vari-
- ants identified in our series, Sn = transmembrane domains, EAG = Ether-à-go-
- 186 go domain, CNBHD = cyclic nucleotide binding homology domain, P = Pore-
- 187 lining loop.
- 188

189 Figure S1. Faces, hands and feet of patients P2, P4, P5, and P7.

- 190 Mild dysmorphic features could be described: long facies with high forehead,
- 191 bilateral epicanthic folds, mild hypertelorism, flat nasal bridge, broad tip of nose,
- 192 flat midface, full lips, small mouth, mild prognathism.
- 193

194 **REFERENCES**

- 1951.Temple, I.K. and M. Baraitser, Severe mental retardation and absent nails of196hallux and pollex. Am J Med Genet, 1991. **41**(2): p. 173-5.
- 197 2. K., Z., *Über anomalien des ektoderms.* Vjschr Zahnheilkd, 1928. **44**: p. 419–34.
- 1983.Laband, P.F., G. Habib, and G.S. Humphreys, Hereditary Gingival199Fibromatosis. Report of an Affected Family with Associated Splenomegaly and200Skeletal and Soft-Tissue Abnormalities. Oral Surg Oral Med Oral Pathol, 1964.201**17**: p. 339-51.
- Bramswig, N.C., C.W. Ockeloen, J.C. Czeschik, A.J. van Essen, R. Pfundt, J.
 Smeitink, B.T. Poll-The, H. Engels, T.M. Strom, D. Wieczorek, T. Kleefstra, and
 H.J. Ludecke, 'Splitting versus lumping': Temple-Baraitser and Zimmermann Laband Syndromes. Hum Genet, 2015. 134(10): p. 1089-97.
- Megarbane, A., R. Al-Ali, N. Choucair, M. Lek, E. Wang, M. Ladjimi, C.M. Rose,
 R. Hobeika, Y. Macary, R. Temanni, P.V. Jithesh, A. Chouchane, K.S. Sastry,
 R. Thomas, S. Tomei, W. Liu, F.M. Marincola, D. MacArthur, and L.
 Chouchane, *Temple-Baraitser Syndrome and Zimmermann-Laband Syndrome:* one clinical entity? BMC Med Genet, 2016. **17**(1): p. 42.
- Simons, C., L.D. Rash, J. Crawford, L. Ma, B. Cristofori-Armstrong, D. Miller, K.
 Ru, G.J. Baillie, Y. Alanay, A. Jacquinet, F.G. Debray, A. Verloes, J. Shen, G.
 Yesil, S. Guler, A. Yuksel, J.G. Cleary, S.M. Grimmond, J. McGaughran, G.F.
 King, M.T. Gabbett, and R.J. Taft, *Mutations in the voltage-gated potassium channel gene KCNH1 cause Temple-Baraitser syndrome and epilepsy.* Nat
 Genet, 2015. 47(1): p. 73-7.

- Kortum, F., V. Caputo, C.K. Bauer, L. Stella, A. Ciolfi, M. Alawi, G.
 Bocchinfuso, E. Flex, S. Paolacci, M.L. Dentici, P. Grammatico, G.C. Korenke,
 V. Leuzzi, D. Mowat, L.D. Nair, T.T. Nguyen, P. Thierry, S.M. White, B.
 Dallapiccola, A. Pizzuti, P.M. Campeau, M. Tartaglia, and K. Kutsche,
 Mutations in KCNH1 and ATP6V1B2 cause Zimmermann-Laband syndrome.
 Nat Genet, 2015. 47(6): p. 661-7.
- Jacquinet, A., M. Gerard, M.T. Gabbett, L. Rausin, J.P. Misson, B. Menten, G.
 Mortier, L. Van Maldergem, A. Verloes, and F.G. Debray, *Temple-Baraitser* syndrome: a rare and possibly unrecognized condition. Am J Med Genet A,
 2010. 152A(9): p. 2322-6.
- 9. Gabbett, M.T., R.C. Clark, and J.M. McGaughran, A second case of severe
 mental retardation and absent nails of hallux and pollex (Temple-Baraitser
 syndrome). Am J Med Genet A, 2008. 146A(4): p. 450-2.
- Shen, J.J., *Two cases of Temple-Baraitser syndrome: natural history and further delineation of the clinical and radiologic phenotypes.* Clin Dysmorphol, 2015. 24(2): p. 55-60.
- 233 11. Van Buggenhout, G.J., H.G. Brunner, J.C. Trommelen, and B.C. Hamel,
 234 *Zimmermann-Laband syndrome in a patient with severe mental retardation.*235 Genet Couns, 1995. 6(4): p. 321-7.
- 236 12. Chodirker, B.N., A.E. Chudley, M.A. Toffler, and M.H. Reed, *Zimmerman-Laband syndrome and profound mental retardation*. Am J Med Genet, 1986.
 238 25(3): p. 543-7.
- 239 13. Castori, M., M. Valiante, G. Pascolini, V. Leuzzi, A. Pizzuti, and P. Grammatico,
 240 *Clinical and genetic study of two patients with Zimmermann-Laband syndrome*241 *and literature review.* Eur J Med Genet, 2013. 56(10): p. 570-6.
- Abo-Dalo, B., M. Roes, S. Canun, M. Delatycki, G. Gillessen-Kaesbach, I.
 Hrytsiuk, C. Jung, B. Kerr, D. Mowat, E. Seemanova, C.E. Steiner, H. Stewart,
 P. Thierry, G. van Buggenhout, S. White, M. Zenker, and K. Kutsche, *No mutation in genes of the WNT signaling pathway in patients with Zimmermann- Laband syndrome.* Clin Dysmorphol, 2008. **17**(3): p. 181-5.
- Mastrangelo, M., I.E. Scheffer, N.C. Bramswig, L.D. Nair, C.T. Myers, M.L.
 Dentici, G.C. Korenke, K. Schoch, P.M. Campeau, S.M. White, V. Shashi, S.
 Kansagra, A.J. Van Essen, and V. Leuzzi, *Epilepsy in KCNH1-related syndromes.* Epileptic Disord, 2016. **18**(2): p. 123-36.
- 16. Fukai, R., H. Saitsu, Y. Tsurusaki, Y. Sakai, K. Haginoya, K. Takahashi, M.W.
 Hubshman, N. Okamoto, M. Nakashima, F. Tanaka, N. Miyake, and N.
 Matsumoto, *De novo KCNH1 mutations in four patients with syndromic developmental delay, hypotonia and seizures.* J Hum Genet, 2016. 61(5): p. 381-7.
- Bauer, C.K., P.E. Schneeberger, F. Kortum, J. Altmuller, F. Santos-Simarro, L.
 Baker, J. Keller-Ramey, S.M. White, P.M. Campeau, K.W. Gripp, and K.
 Kutsche, *Gain-of-Function Mutations in KCNN3 Encoding the Small- Conductance Ca(2+)-Activated K(+) Channel SK3 Cause Zimmermann-Laband*Syndrome. Am J Hum Genet, 2019. **104**(6): p. 1139-1157.
- 18. Guglielmi, F., E. Staderini, F. lavarone, L. Di Tonno, and P. Gallenzi, *Zimmermann-Laband-1 Syndrome: Clinical, Histological, and Proteomic Findings of a 3-Year-Old Patient with Hereditary Gingival Fibromatosis.*Biomedicines, 2019. 7(3).
- 26519.Yesil, G., S. Guler, A. Yuksel, and Y. Alanay, Report of a patient with Temple-266Baraitser syndrome. Am J Med Genet A, 2014. **164A**(3): p. 848-51.

clic expression of the voltage-gated
sembly of the primary cilium. EMBO
al insights into the mechanisms of
18. 150 (2): p. 225-244.



A. cDNA level: in bold, the novel variants reported here.

B. protein level: Legend: Stars representing variants identified, In blue = variants identified in our series, Sn = transmembrane domains, EAG = Ether-à-go-go domain, CNBHD = cyclic nucleotide

binding homology domain, P = Pore-lining loop.

278

Supplementary methods

279

The DNA of patient 1 (P1) was analyzed on a 116 epilepsy gene panel. Nextgeneration sequencing was performed using a SureSelectXT Custom 12-24Mb library (Agilent Technologies) on a ion proton platform (Thermo Fisher). The results were analyzed using the Torrent suite (Thermo Fisher) for alignment and variant calling. To filter and establish coverage and variant analysis, we used the Varaft annotation and filtration system (Desvignes et al., Nucleic Acids Res. 2018 Jul 2; 46: W545–W553.doi: 10.1093/nar/gky471)

287

The DNA of patient 4 (P4) and patient 5 (P5) was analyzed on a 90 epilepsy gene panel. A library of all coding exons and intron-exon boundaries was prepared using a SeqCap EZ Library (Roche-NimbleGen) following the manufacturer's instructions. Genepanel sequencing was performed on a MiSeq (Illumina Inc). Sequence alignment was performed by Genodiag with BWA 0.7.12, picard-tools-1.121, GATK-3.5 (Indel realignment, base recalibration). GATK-3.5 (HaplotypeCaller) and SNPEff-4.2 were used for variant calling and annotation, adding information from gnomAD, ClinVar and HGMD.

295

The DNA of patients 2 (P2), 3 (P3), 6 (P6) and 7 (P7) and their parents was studied by trio-WES. A library of all coding exons and intron-exon boundaries was prepared using a SeqCap EZ MedExome (Roche-NimbleGen) following the manufacturer's instructions. Whole Exome sequencing was performed on the Illumina NextSeq 500 platform. <u>P2, P3 and P6:</u> Raw reads were mapped to the human genome reference-build hg19 using the Burrows Wheeler Aligner (BWA MEM v0.717) alignment algorithm. The re-

sulting binary alignment/map (BAM) files were further processed by Genome Analysis
 Tool Kit HaplotypeCaller (GATK HC v3.8). The VCF files were then annotated on
 Snpeff version 4.3T. Only coding non synonymous and splicing variants were consid-

305 ered. Variant prioritization was conducted thanks to the transmission mode (*de novo*,
306 autosomal recessive and X-linked), and the frequency of the variants in the gnomAD
307 database.

308 P7: Fastq files were aligned to human genome hg19 with bwa mem (v0.7.3). We then 309 called SNVs and INDELs following GATKs best practices (v3.4). We achieved an aver-310 age mean target coverage of 113X. Variants were annotated using ANNOVAR and 311 filtered with in-house scripts to keep variants with at least 9 reads and with a variant 312 read frequency over 20 percent impacting exonic sequences or splice sites (+/- 10bp 313 from the junction) and with an allele frequency <0.5% in 1000 genomes, genome ag-314 gregation database (gnomAD, 123,136 exomes and 15,496 whole genome sequences; 315 accessed on 11/10/2018) and in a local database. The possible functional impact of 316 amino-acid changes was predicted by SIFT (Sorting Intolerant from Tolerant), Poly-317 Phen-2 hvar and CADD score (Combined Annotation Dependent Depletion). The Ala-318 mut software (Interactive biosoftware) was used to study retained variant sites.



Figure S1. Faces, hands and feet of patients P2, P4, P5, and P7. Mild dysmorphic features could be described: long facies with high forehead, bilateral epicanthic folds, mild hypertelorism, flat nasal bridge, broad tip of nose, flat midface, full lips, small mouth, mild prognathism.